Package: mSigTools (via r-universe)

September 7, 2024

Type Package

Title Mutational Signature Analysis Tools

Version 1.0.7

Description Utility functions for mutational signature analysis as described in Alexandrov, L. B. (2020) <doi:10.1038/s41586-020-1943-3>. This package provides two groups of functions. One is for dealing with mutational signature ``exposures'' (i.e. the counts of mutations in a sample that are due to each mutational signature). The other group of functions is for matching or comparing sets of mutational signatures. 'mSigTools' stands for mutational Signature analysis Tools.

License GPL-3

URL https://github.com/Rozen-Lab/mSigTools

BugReports https://github.com/Rozen-Lab/mSigTools/issues

Encoding UTF-8

Language en-US

LazyData true

RoxygenNote 7.2.3

Imports clue, philentropy, quadprog, sets

Suggests cosmicsig, ICAMS, spelling, testthat (>= 3.0.0),

Config/testthat/edition 3

Repository https://rozen-lab.r-universe.dev

RemoteUrl https://github.com/rozen-lab/msigtools

RemoteRef HEAD

RemoteSha 48994a13b8380388556768dab0d3e06602516c6b

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find_best_reconstruction_QP

Find "best" reconstruction of a target signature or spectrum from a set of signatures.

Description

Find "best" reconstruction of a target signature or spectrum from a set of signatures.

Usage

```
find_best_reconstruction_QP(
  target.sig,
  sig.universe,
  max.subset.size = NULL,
  method = "cosine",
  trim.less.than = 1e-10
)
```

Arguments

target.sig   The signature or spectrum to reconstruct; a non-negative numeric vector or 1-column matrix-like object.

sig.universe   The universe of signatures from which to reconstruct target.sig; a non-negative numeric matrix-like object with nrow(sig.universe) == length(target.sig). The sums of each column must be 1. Must not contain duplicate columns or have other non-unique quadratic programming solutions (not checked, but will generate an error from solve.QP in package quadprog).

max.subset.size

                  Maximum number of signatures to use to reconstruct target.sig.

method	As in [dist_one_one](#) in package philentropy, and used only to find the final "best" reconstruction. The optimized exposures from which to selected the "best" reconstruction are calculated using [optimize_exposure_QP](#), which uses [solve.QP](#) in package quadprog.
trim.less.than	After optimizing exposures with [optimize_exposure_QP](#), remove exposures less than trim.less.than and then re-optimize.

Details

This function should be fast if you do not specify max.subset.size, but it will be combinatorially slow if max.subset.size is large and trim.less.than is small or negative. So do not do that. If max.subset.size is NULL, then the function just uses [optimize_exposure_QP](#). and then excludes exposures < trim.less.than, and then re-runs [optimize_exposure_QP](#). Otherwise, after excluding exposures < trim.less.than, then the function runs [optimize_exposure_QP](#) on subsets of signatures of size <= max.subset.size, removes exposures < trim.less.than, reruns [optimize_exposure_QP](#), calculates the reconstruction and similarity between the reconstruction and the target.sig and returns the information for the exposures that have the greatest similarity.

Value

A list with elements:

- optimized.exposure A numerical vector of the exposures that give the "best" reconstruction. This vector is empty if there is an error.

- similarity The similarity between the reconstruction (see below) and target.sig according to the distance or similarity provided by the method argument.

- method The value specified for the method argument, or an error message if optimize.exposure is empty.

- reconstruction The reconstruction of target.sig according to optimized.exposure.

Examples

```
set.seed(888)
sig.u <-
  do.call(
    cbind,
    lapply(1:6, function(x) {
      col <- runif(n = 96)
      col / sum(col)
    })
  )
rr <- find_best_reconstruction_QP(
  target.sig = sig.u[, 1, drop = FALSE],
  sig.universe = sig.u[, 2:6]
)
names(rr)
rr$optimized.exposure
rr$similarity
rr <- find_best_reconstruction_QP(
  target.sig = sig.u[, 1, drop = FALSE],
```

```
    sig.universe = sig.u[, 2:6],
    max.subset.size = 3
)
rr$optimized.exposure
rr$similarity
```

---

match_two_sig_sets   *Find an optimal matching between two sets of signatures subject to a*
                            maximum distance.

Description

Find an optimal matching between two sets of signatures subject to a maximum distance.

Usage

```
match_two_sig_sets(
  x1,
  x2,
  method = "cosine",
  convert.sim.to.dist = function(x) {
      return(1 - x)
  },
  cutoff = 0.9
)
```

Arguments

x1	A numerical-matrix-like object with columns as signatures.
x2	A numerical-matrix-like object with columns as signatures. Needs to have the same number of rows as x1.
method	As for the distance function in package philenropy.
convert.sim.to.dist	
	If method specifies a similarity rather than a distance, use this function to convert the similarity to a distance.
cutoff	A maximum distance or minimum similarity over which to pair signatures between x1 and x2.

Details

Match signatures between x1 and x2 using the function solve_LSAP, which uses the "Hungarian" (a.k.a "Kuhn–Munkres") algorithm https://en.wikipedia.org/wiki/Hungarian_algorithm, which optimizes the total cost associated with the links between nodes. This function generates a distance matrix between the two sets of signatures using method and, if necessary, convert.sim.to.dist. It then sets distances > cutoff to very large values and then applies solve_LSAP to the resulting matrix to compute a matching between x1 and x2 that minimizes the sum of the distances.

Value

A list with the elements

- table Table of extracted signatures that matched a reference signature. Each row contains the extracted signature name, the reference signature name, and the distance of the match.
- orig.matrix The matrix of numeric distances between x1 and x2.
- modified.matrix The argument orig.matrix with distances > cutoff changed to very large values.

Examples

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.6, 0.4), nrow = 2)
colnames(ex.sigs) <- c("ex1", "ex2", "ex3")
ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2)
colnames(ref.sigs) <- c("ref1", "ref2")
match_two_sig_sets(ex.sigs, ref.sigs, cutoff = .9)
```

---

optimize_exposure_QP   *Quadratic programming optimization of signature activities*

Description

Quadratic programming optimization of signature activities

Usage

```
optimize_exposure_QP(spectrum, signatures)
```

Arguments

spectrum   Mutational signature or mutational spectrum as a numeric vector or single column data frame or matrix.

signatures   Matrix or data frame of signatures from which to reconstruct spectrum. Rows are mutation types and columns are signatures. Should have column names for interpretable results. Cannot be a vector because the column names are needed.

Details

Code adapted from SignatureEstimation::decomposeQP and uses solve.QP in package quadprog.

Value

A vector of exposures with names being the colnames from signatures.

Examples

```
usigs <- matrix(c(0.2, 0.7, 0.1,
                  0.3, 0.6, 0.1,
                  0.1, 0.1, 0.8), nrow = 3)
colnames(usigs) <- c("u1", "u2", "u3")
tsig <- matrix(c(0.25, 0.65, 0.1), nrow = 3)
optimize_exposure_QP(tsig, usigs)
```

---

plot_exposure   *Plot exposures in multiple plots, with each plot showing exposures for*
                         a manageable number of samples.

Description

Plot exposures in multiple plots, with each plot showing exposures for a manageable number of
samples.

Usage

```
plot_exposure(
  exposure,
  samples.per.line = 30,
  plot.proportion = FALSE,
  xlim = NULL,
  ylim = NULL,
  legend.x = NULL,
  legend.y = NULL,
  cex.legend = 0.9,
  cex.yaxis = 1,
  cex.xaxis = NULL,
  plot.sample.names = TRUE,
  yaxis.labels = NULL,
  ...
)
```

Arguments

exposure   Exposures as a numerical `matrix` (or `data.frame`) with signatures in rows and
                    samples in columns. Rownames are taken as the signature names and column
                    names are taken as the sample IDs. If you want `exposure` sorted from largest to
                    smallest, use `sort_exposure`. Do not use column names that start with multiple
                    underscores. The exposures will often be mutation counts, but could also be e.g.
                    mutations per megabase.

samples.per.line
                    Number of samples to show in each plot.

plot.proportion
                    Plot exposure proportions rather than counts.

xlim, ylim	Limits for the x and y axis. If NULL(default), the function tries to do something reasonable.
legend.x, legend.y	
	The x and y co-ordinates to be used to position the legend.
cex.legend	A numerical value giving the amount by which legend plotting text and symbols should be magnified relative to the default.
cex.yaxis	A numerical value giving the amount by which y axis values should be magnified relative to the default.
cex.xaxis	A numerical value giving the amount by which x axis values should be magnified relative to the default. If NULL(default), the function tries to do something reasonable.
plot.sample.names	
	Whether to plot sample names below the x axis. Default is TRUE. Ignored if there are no column names on exposure.
yaxis.labels	User defined y axis labels to be plotted. If NULL(default), the function tries to do something reasonable.
...	Other arguments passed to [barplot](). If ylab is not included, it defaults to a value depending on plot.proportion. If col is not supplied the function tries to do something reasonable.

Value

An **invisible** list. The first element is a logical value indicating whether the plot is successful. The second element is a numeric vector giving the coordinates of the bar x-axis midpoints drawn, useful for adding to the graph.

Examples

```
file <- system.file("extdata",
  "Liver-HCC.exposure.csv",
  package = "mSigTools"
)
exposure <- read_exposure(file)
old.par <- par(mar = c(8, 5, 1, 1))
plot_exposure(exposure[, 1:30],
  main = "Liver-HCC exposure", cex.yaxis = 0.8,
  plot.proportion = TRUE
)
par(old.par)
```

---

plot_exposure_to_pdf	*Plot exposures in multiple plots to a single PDF file, with each plot showing exposures for a manageable number of samples.*

Description

Plot exposures in multiple plots to a single PDF file, with each plot showing exposures for a manageable number of samples.

Usage

```
plot_exposure_to_pdf(
  exposure,
  file,
  mfrow = c(2, 1),
  mar = c(6, 4, 3, 2),
  oma = c(3, 2, 0, 2),
  samples.per.line = 30,
  plot.proportion = FALSE,
  xlim = NULL,
  ylim = NULL,
  legend.x = NULL,
  legend.y = NULL,
  cex.legend = 0.9,
  cex.yaxis = 1,
  cex.xaxis = NULL,
  plot.sample.names = TRUE,
  yaxis.labels = NULL,
  width = 8.2677,
  height = 11.6929,
  ...
)
```

Arguments

exposure	Exposures as a numerical `matrix` (or `data.frame`) with signatures in rows and samples in columns. Rownames are taken as the signature names and column names are taken as the sample IDs. If you want exposure sorted from largest to smallest, use [`sort_exposure`](). Do not use column names that start with multiple underscores. The exposures will often be mutation counts, but could also be e.g. mutations per megabase.
file	The name of the PDF file to be produced.
mfrow	A vector of the form `c(nr, nc)`. Subsequent figures will be drawn in an `nr`-by-`nc` array on the device by rows.
mar	A numerical vector of the form `c(bottom,left, top, right)` which gives the number of lines of margin to be specified on the four sides of the plot.
oma	A vector of the form `c(bottom, left, top,right)` giving the size of the outer margins in lines of text.
samples.per.line	
	Number of samples to show in each plot.
plot.proportion	
	Plot exposure proportions rather than counts.
xlim, ylim	Limits for the x and y axis. If `NULL`(default), the function tries to do something reasonable.
legend.x, legend.y	
	The x and y co-ordinates to be used to position the legend.

cex.legend	A numerical value giving the amount by which legend plotting text and symbols should be magnified relative to the default.
cex.yaxis	A numerical value giving the amount by which y axis values should be magnified relative to the default.
cex.xaxis	A numerical value giving the amount by which x axis values should be magnified relative to the default. If NULL(default), the function tries to do something reasonable.
plot.sample.names	
	Whether to plot sample names below the x axis. Default is TRUE. Ignored if there are no column names on exposure.
yaxis.labels	User defined y axis labels to be plotted. If NULL(default), the function tries to do something reasonable.
width, height	The width and height of the graphics region in inches.
...	Other arguments passed to [barplot](). If ylab is not included, it defaults to a value depending on plot.proportion. If col is not supplied the function tries to do something reasonable.

Value

An **invisible** list. The first element is a logical value indicating whether the plot is successful. The second element is a numeric vector giving the coordinates of the bar x-axis midpoints drawn, useful for adding to the graph.

Examples

```
file <- system.file("extdata",
  "Liver-HCC.exposure.csv",
  package = "mSigTools"
)
exposure <- read_exposure(file)
plot_exposure_to_pdf(exposure,
  file = file.path(tempdir(), "Liver-HCC.exposure.pdf"),
  cex.yaxis = 0.8, plot.proportion = TRUE
)
```

---

read_exposure   *Read an exposure matrix from a file.*

Description

Read an exposure matrix from a file.

Usage

```
read_exposure(file, check.names = FALSE)
```

Arguments

`file`	File path to a CSV file containing an exposure matrix, i.e. the numbers of mutations due to each mutational signature. Each row corresponds to a mutational signature an each column corresponds to a tumor or other biological sample.
`check.names`	Passed to `read.csv`. **IMPORTANT**: If `TRUE` this will replace the double colon in identifiers of the form <tumor_type>::<sample_id> with two periods (i.e. <tumor_type>..<sample_id>. If `check.names` is true, generate a warning if double colons were present.

Value

Numerical matrix of exposures, with the same shape as the contents of `file`.

Examples

```
file <- system.file("extdata",
  "Liver-HCC.exposure.csv",
  package = "mSigTools"
)
exposure <- read_exposure(file)
```

---

sig_dist_matrix	*Compute a matrix of distances / similarities between two sets of signatures.*

Description

Compute a matrix of distances / similarities between two sets of signatures.

Usage

```
sig_dist_matrix(x1, x2, method = "cosine")
```

Arguments

`x1`	The first set of signatures (a numerical matrix-like object in which each column is a signature).
`x2`	The second set of signatures, similar data type to `x1`, and must have the same number of rows as `x1`.
`method`	As for the distance function in package `philenropy`.

Value

A numeric matrix with dimensions `ncol(x1)` X `ncol(x2)`. Each element represents the distance or similarity (depending on `method`) between a column in `x1` and a column in `x2`.

Examples

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.4, 0.6), nrow = 2)
colnames(ex.sigs) <- c("ex1", "ex2", "ex3")
ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2)
colnames(ref.sigs) <- c("ref1", "ref2")
sig_dist_matrix(ex.sigs, ref.sigs)
```

---

sort_exposure	*Sort columns of an exposure matrix based on the number of mutations in each sample (column).*

Description

Sort columns of an exposure matrix based on the number of mutations in each sample (column).

Usage

```
sort_exposure(exposure, decreasing = TRUE)
```

Arguments

exposure	Exposures as a numerical matrix (or data.frame) with signatures in rows and samples in columns. Rownames are taken as the signature names and column names are taken as the sample IDs.
decreasing	If TRUE, sort from largest to smallest.

Value

The original exposure with columns sorted.

Examples

```
file <- system.file("extdata",
  "Liver-HCC.exposure.csv",
  package = "mSigTools"
)
exposure <- read_exposure(file)
exposure.sorted <- sort_exposure(exposure)
```

TP_FP_FN_avg_sim   *Find best matches (by cosine similarity) of a set of mutational signatures to a set of reference mutational signatures.*

Description

Find best matches (by cosine similarity) of a set of mutational signatures to a set of reference mutational signatures.

Usage

```
TP_FP_FN_avg_sim(extracted.sigs, reference.sigs, similarity.cutoff = 0.9)
```

Arguments

extracted.sigs   Mutational signatures discovered by some analysis. A numerical-matrix-like object with columns as signatures.

reference.sigs   A numerical-matrix-like object with columns as signatures. This matrix should contain the reference mutational signatures. For example, these might be from a synthetic data set or they could be from reference set of signatures, such as the signatures at the COSMIC mutational signatures web site. See CRAN package cosmicsig.

similarity.cutoff
                 A signature in reference.sigs must be matched by >= similarity.cutoff by a signature in extracted.sigs to be considered detected.

Details

Match signatures in extracted.sigs to signatures in reference.sigs using match_two_sig_sets based on cosine similarity.

Value

A list with the elements

- TP The number of true positive extracted signatures.
- FP The number of false positive extracted signatures.
- FN The number of false negative reference signatures.
- avg.cos.sim The average cosine similarity of true positives to their matching reference signatures.
- table A data.frame of extracted signatures that matched a reference signature. Each row contains the extracted signature name, the reference signature name, and the cosine similarity of the match.
- sim.matrix The numeric distance or similarity matrix between extracted.sigs and reference.sigs as returned from sig_dist_matrix.

- `unmatched.ex.sigs` The identifiers of the extracted signatures that did not match a reference signature.
- `unmatched.ref.sigs` The identifiers of the reference signatures that did not match an extracted signature.

Examples

```
ex.sigs <- matrix(c(0.2, 0.8, 0.3, 0.7, 0.6, 0.4), nrow = 2)
colnames(ex.sigs) <- c("ex1", "ex2", "ex3")
ref.sigs <- matrix(c(0.21, 0.79, 0.19, 0.81), nrow = 2)
colnames(ref.sigs) <- c("ref1", "ref2")
TP_FP_FN_avg_sim(
  extracted.sigs = ex.sigs,
  reference.sigs = ref.sigs,
  similarity.cutoff = .9
)
```

---

write_exposure   *Write an exposure matrix to a file.*

Description

Write an exposure matrix to a file.

Usage

```
write_exposure(exposure, file, row.names = TRUE)
```

Arguments

exposure	Exposures as a numerical matrix (or data.frame) with signatures in rows and samples in columns. Rownames are taken as the signature names and column names are taken as the sample IDs.
file	File to which to write the exposure matrix (as a CSV file).
row.names	Either a logical value indicating whether the row names of exposure are to be written along with exposure, or a character vector of row names to be written.

Value

No return value, called for side effects.

Examples

```
file <- system.file("extdata",
  "Liver-HCC.exposure.csv",
  package = "mSigTools"
)
exposure <- read_exposure(file)
write_exposure(exposure, file = file.path(tempdir(), "Liver-HCC.exposure.csv"))
```

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